

Introduction to Statistical Computing in Clinical Research

BIOSTAT 212, Summer 2016



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Outline today

◇ Course details

◇ Lecture 1

- Why STATA?
- Orientation to STATA
- Immediate commands

◇ Mini-lecture: Sample size calculations in STATA

◇ Lecture 2

- Opening data sets
- Workflow: .do files and log files

Introductions

- ◊ Who are you ?
- ◊ Teaching Assistants
 - Biftu Mengesha
 - Swaroop Mistry
 - Anjana Sharma
 - Yiwey Shieh
 - Qurratul Warsi

Acknowledgements

◊ Mark Pletcher

Course Objectives

- ◇ Introduce you to using STATA for:
 - Data management
 - Basic statistical and epidemiologic analysis
 - Turning raw data into presentable tables, figures and other research products
- ◇ Prepare you for fall courses
- ◇ Start analyzing your own data

Course Objectives

◊ NOT statistical theory

- Biostat200 in the fall is the first biostats class
- I'm an epidemiologist, not a statistician

Course details

- ◇ 1 Unit Course
- ◇ 7 Lectures
 - Tuesdays 1:15 – 2:45
- ◇ 6 Labs
 - Tuesdays 3:00 – 4:00 (except for today)
- ◇ 5 Assignments
- ◇ Additional online Lab Session/Office Hours
 - Wednesdays 10:00-11:00

Course Details

◇ Course Website

- <https://courses.ucsf.edu/course/view.php?id=3039>

◇ Recorded Lectures

- Audio + video of lecturer + video of screen
- Available same day for viewing
- Links posted on website syllabus

Course details

◇ Online forum

- **Post all questions here!** (no email please)
 - TA's will staff Forum, reply by end of day on weekdays
- Before you post, see if it's already there and answered
- Look at previous year's forums for useful information
- Settings for email notifications
 - <https://learningtech.library.ucsf.edu/-/?s=forum+email+notifications>

Course details

◇ Course Requirements

- Hand in all 5 Assignments (even if late)
- Satisfactory Final Project

Course details

◊ Grading (not relevant for all students)

- Letter grades: Standard cutoffs

○ 90-100%	A
○ 80-89%	B
○ 70-79%	C
○ 60-69%	D
○ <60% or Course Requirements not met:	F

- Satisfactory/Unsatisfactory

○ >80%	Satisfactory
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- TICR Grading policy

- http://www.epibiostat.ucsf.edu/courses/masters/grading_policy.html

Lecture Topics

1. Introduction, orientation and immediate commands
2. .do files, .log files, and workflow
3. Importing data, exploring data and generating variables
4. Cleaning data
5. Statistical analysis
6. Epidemiologic analysis
7. Tables and figures

Mini-lectures (given by TAs)

1. Sample size calculations
2. Reshaping data
3. Dealing with dates
4. Formulas in excel
5. Loops
6. Add-on packages

Lab Sessions

- ◇ Most important element of the course!
- ◇ Time to do your assignment and ask for help

Assignments

◇ 5 Assignments

- ◇ Assignments are due 1 week after they are assigned prior to lecture.
 - Please upload to the course website. **By 1:00 PM**
- ◇ Assignments turned in late (less than 1 week) will receive only half credit; after that, no points will be awarded.
- ◇ ALL labs must be turned in to pass the class (even if no points are awarded).

Final Project

- ◇ Create a table and a figure using your own data, document analysis using Stata.
- ◇ Due **September 13 at 9 PM**
- ◇ 30 points docked for each 1 day late.
- ◇ See 1-page description and example in syllabus
- ◇ Start looking for data!

Lecture 1: Introduction to STATA

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Many Software Packages

- ◇ SAS
- ◇ R
- ◇ SPSS
- ◇ Epi-Info
- ◇ STATA
- ◇

Why use STATA?

- ◇ Quick start, user friendly, but full-featured and powerful
- ◇ Immediate results
- ◇ Command line and Menu-driven option
- ◇ Good graphics
- ◇ Log and .do files
- ◇ Good manuals, help menu, customer service
- ◇ UCSF

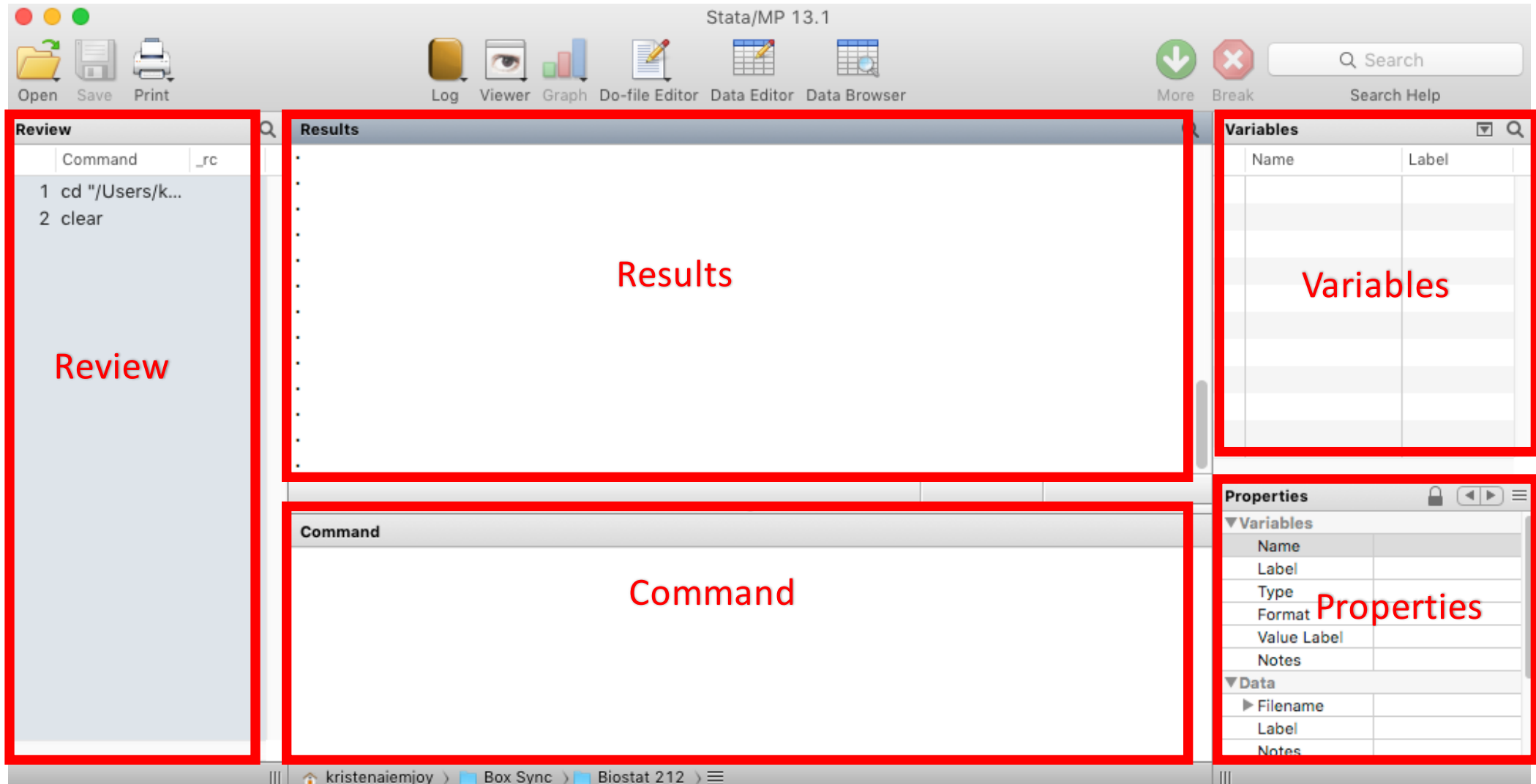
Why not STATA?

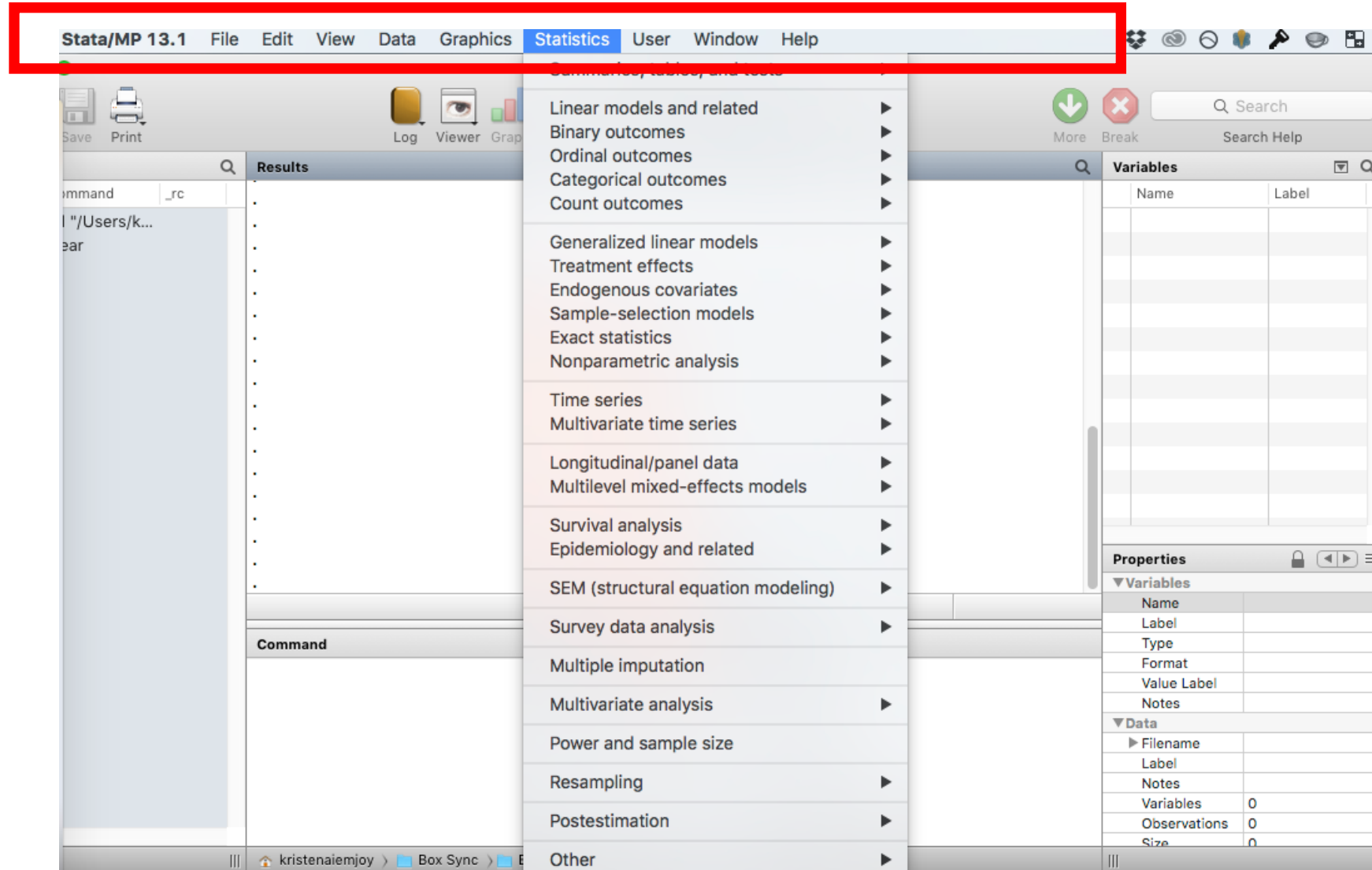
- ◇ \$\$\$
- ◇ R and Epi-info are free
- ◇ Programming easier in R
- ◇ Graphics can be better in R
- ◇ In STATA, you can only have one data frame open at a time

STATA vs. other software packages

Features	Stata	SPSS	SAS	R
Learning curve	Moderate	Gradual/flat	Pretty steep	Pretty steep
User interface	Programming/point-and-click	Mostly point-and-click	Programming	Programming
Data manipulation	Very strong	Moderate	Very strong	Very strong
Data analysis	Powerful	Powerful	Powerful/versatile	Powerful/versatile
Graphics	Very good	Very good	Good	Excellent
Cost	Affordable (perpetual licenses, renew only when upgrade)	Expensive (but not need to renew until upgrade, long term licenses)	Expensive (yearly renewal)	Open source

Windows





Menu versus Command Line

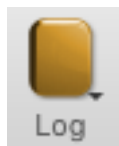
◇ Menu advantages

- Browse for commands you don't know already
- See the options for each command in dialog boxes
- Good way to learn syntax for complex commands

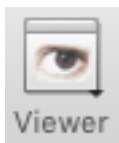
◇ Command line advantages

- MUCH faster
- ONLY way to write “do” files
 - Document and repeat analyses

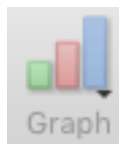
Buttons



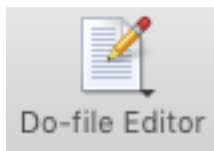
Log Begin/Close/Suspend/Resume Starts or closes a file to record results



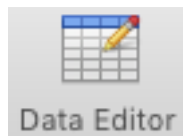
Viewer Opens a help window



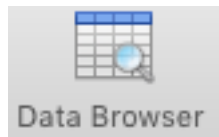
Graph Brings Graph Window to Front



Do-file Editor Opens a new .do (command) file



Data Editor Opens data window and allows edits



Data Browser Opens data window to browse but not edit

Break and More

When there are more results than will fit in your result window use:



Or the

Space

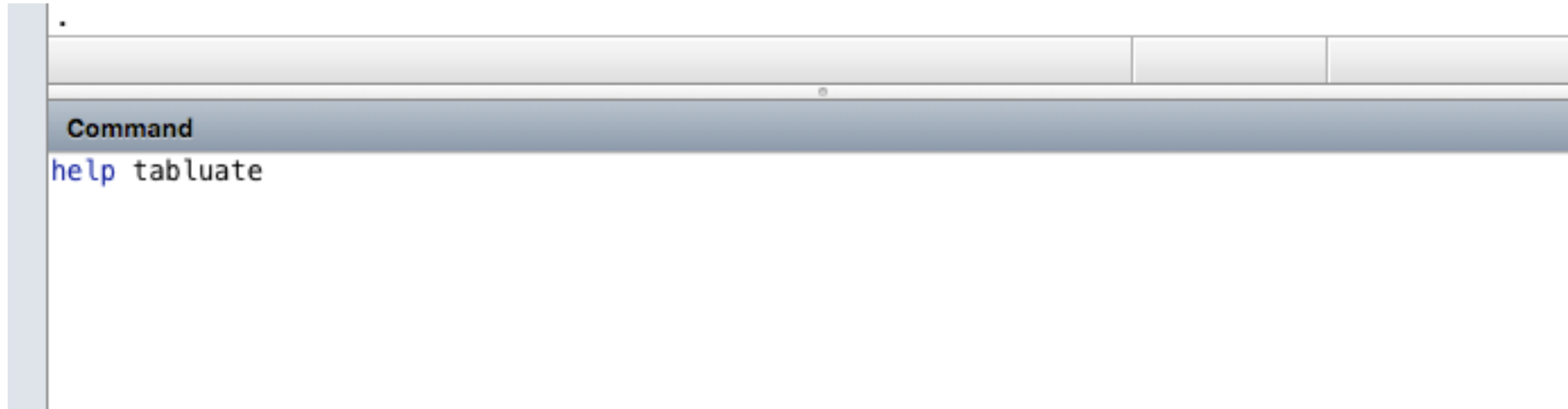
To see more



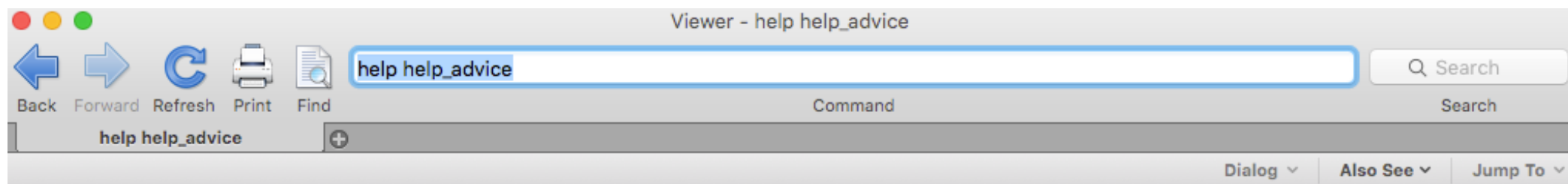
To stop

*note: you can also use the command `set more on`
`set more off`

Help!



Help!



Advice on finding help

Let's say you are trying to find out how to do something. With over 2,000 help files and 11,000 pages of PDF documentation, we have probably explained how to do whatever you want. The documentation is filled with worked examples that you can run on supplied datasets. Whatever your question, try the following first:

1. Select **Help** from the Stata menu and click on **Search...**
2. Type some keywords about the topic that interests you, say, "logistic regression".
3. Look through the resulting list of available resources, including help files, FAQs, Stata Journal articles, and other resources.
4. Select the resource whose description looks most helpful. Usually, this description will be a help file and will include "(help <xyz>)", or, as in our example, perhaps "(help [logistic](#))". Click on the blue link "logistic".
5. Let's assume you have selected the "(help [logistic](#))" entry. You are probably not interested in the syntax at the top of the file, but you would like to see some examples. Select **Jump To** from the Viewer menu (in the top right corner of the help file now on your screen), and click on **Examples**. You will be taken to example commands that you can run on example datasets. Simply cut and paste those commands into Stata to see the results.
6. If you are new to the **logistic** command and want both an overview and worked examples with discussion, from the **Also See** menu, click on [\[R\] logistic](#) with the PDF icon. Or at the top of the help file, click on the blue title of the

Help!

[All](#) [Images](#) [Videos](#) [News](#) [Shopping](#) [More ▾](#) [Search tools](#)

About 125,000 results (0.50 seconds)

[Stata | FAQ: Creating percent summary variables](#)
[www.stata.com](#) ▸ [Resources & support](#) ▾ Stata ▾
How can I create variables containing **percent** summaries? ... sysuse auto (1978 Automobile Data) .
`tabulate rep78 foreign, row +-----+ ....` For **example**,

[\[PDF\] tabulate twoway - Stata](#)
[www.stata.com/manuals13/rtabulatetwoway.pdf](#) ▾ Stata ▾
Unix(console) users can instead use the `set linesize` **command** to take advantage of `tabulate` can
calculate column **percentages** and cell **percentages**, as well.

[st: RE: Percentages in cross-tabulation - Stata](#)
[www.stata.com/statalist/archive/2002-09/msg00432.html](#) ▾ Stata ▾
Sep 24, 2002 - The > problem is how do I get column **percentages** of income > without transferring the
tabulation results to XL? See the on-line **help**. . `tab sex ...`

STATA syntax

1. I will use `Courier font` for commands
2. Words that are NOT meant to be typed in but instead are descriptive placeholders will be `blue`.
3. Parts of the command that are optional will be placed in `[brackets]`.
4. Shortcuts for commands are indicated by underlining
 - summarize

STATA syntax

`varlist`

1 or more variables

`varname`

One variable

`var1, var2`

Exactly 2 variables

`#`

A number (in immediate commands)
aka `numlist` in stata help files

STATA Syntax

tabulate `var1 var2`, [row]

`tab male smoke, row`

male	smoke		Total
	0	1	
0	2,723 65.00	1,466 35.00	4,189 100.00
1	3,539 57.24	2,644 42.76	6,183 100.00
Total	6,262 60.37	4,110 39.63	10,372 100.00

STATA Syntax

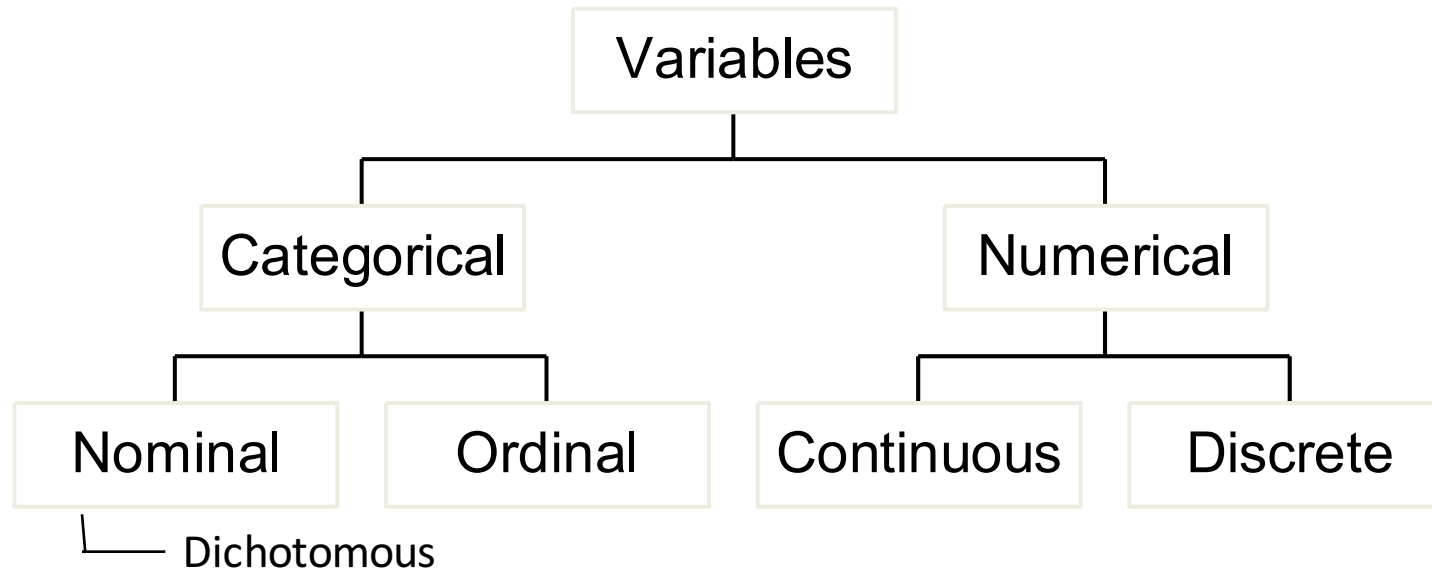
◇ STATA is case sensitive

- Commands are in lower case
- If your variables have upper case letters, you need to remember this when typing them into the command line
 - Often easier to use all lower case

Types of Files

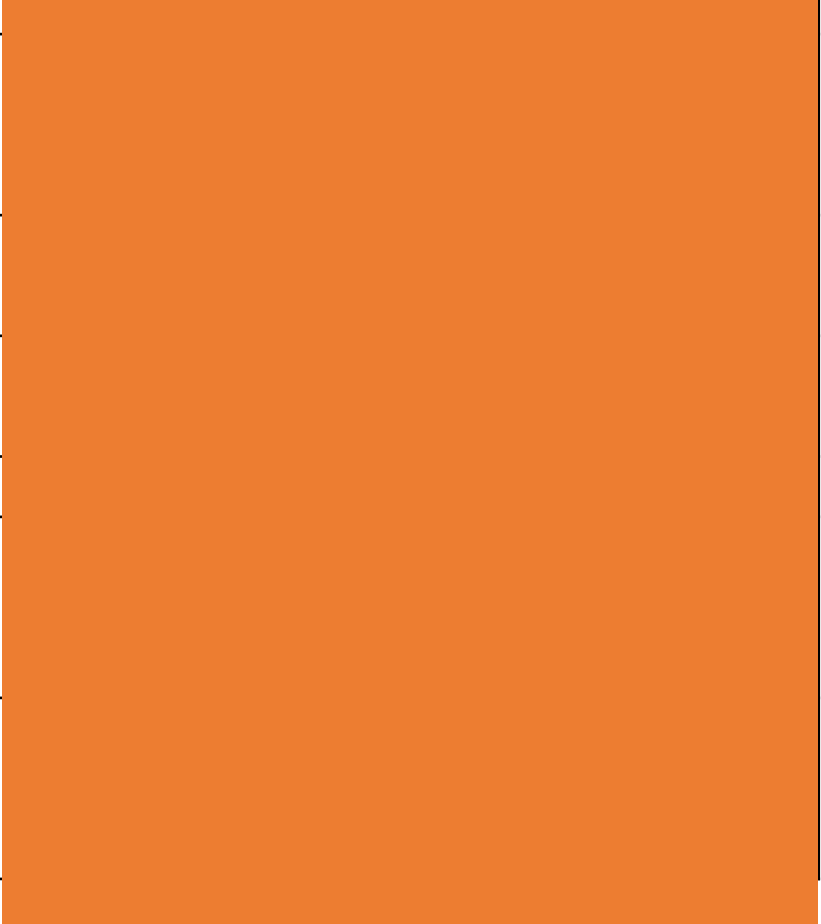
.dta	data files
.do	command files
.ado	programs
.hlp	help files
.gph	graphs
.dct	dictionary files

**Preview: types of Variables



Types of Variables

Categorical	
1. Nominal	No order
→ 1a. Dichotomous	No order, only 2 categories
2. Ordinal	Ordered
Numerical	
1. Discrete	Can be counted
2. Continuous	Can take any value within a given range



A quick note on binary

◊ 0 No

◊ 1 Yes

STATA Variable Storage Types

str1	string 1 character long	words
str x	string x characters long	
strL	string up to 2 billion characters long	
byte	dichotomous	true/false
int	integer values between -32,767 and 32,740	numbers
long	integer values between -2,147,483,647 and 2,147,483,620	
float	real numbers (i.e. numbers with decimal places) with 8 digits of accuracy	
double	real numbers (i.e. numbers with decimal places) with 16 digits of accuracy	
Missing	Missing data	no data

Variable storage types

obstime	anycac	caccat	agecat	male	age_ge60	sex	haircolor_~e
1125	0	0	<30	1	0	male	blond
1125	0	0	<30	0	0	female	black
1064	0	0	<30	0	0	female	black
1064	0	0	<30	0	0	female	black
913	0	0	<30	1	0	male	black
852	0	0	<30	0	0	female	blond
699	1	1-99	<30	1	0	male	black
1674	0	0	<30	1	0	male	blond
1460	0	0	<30	0	0	female	black
1460	0	0	<30	0	0	female	red
1368	0	0	<30	1	0	male	brown
1156	0	0	<30	1	0	male	blond

Blue Categorical with label
Black Numeric or categorical with no label
Red String

Demo 1

Useful keyboard shortcuts



Describe data



+



Open a .do file



+



Execute command from
.do file (highlight text)



scroll through previous commands

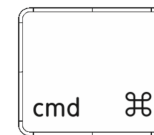


Autocomplete variable name after typing part of it

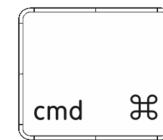
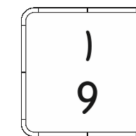
Mac



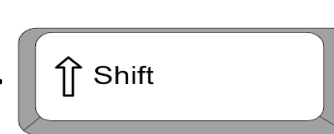
+



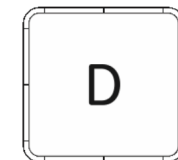
+



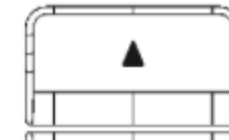
+



+



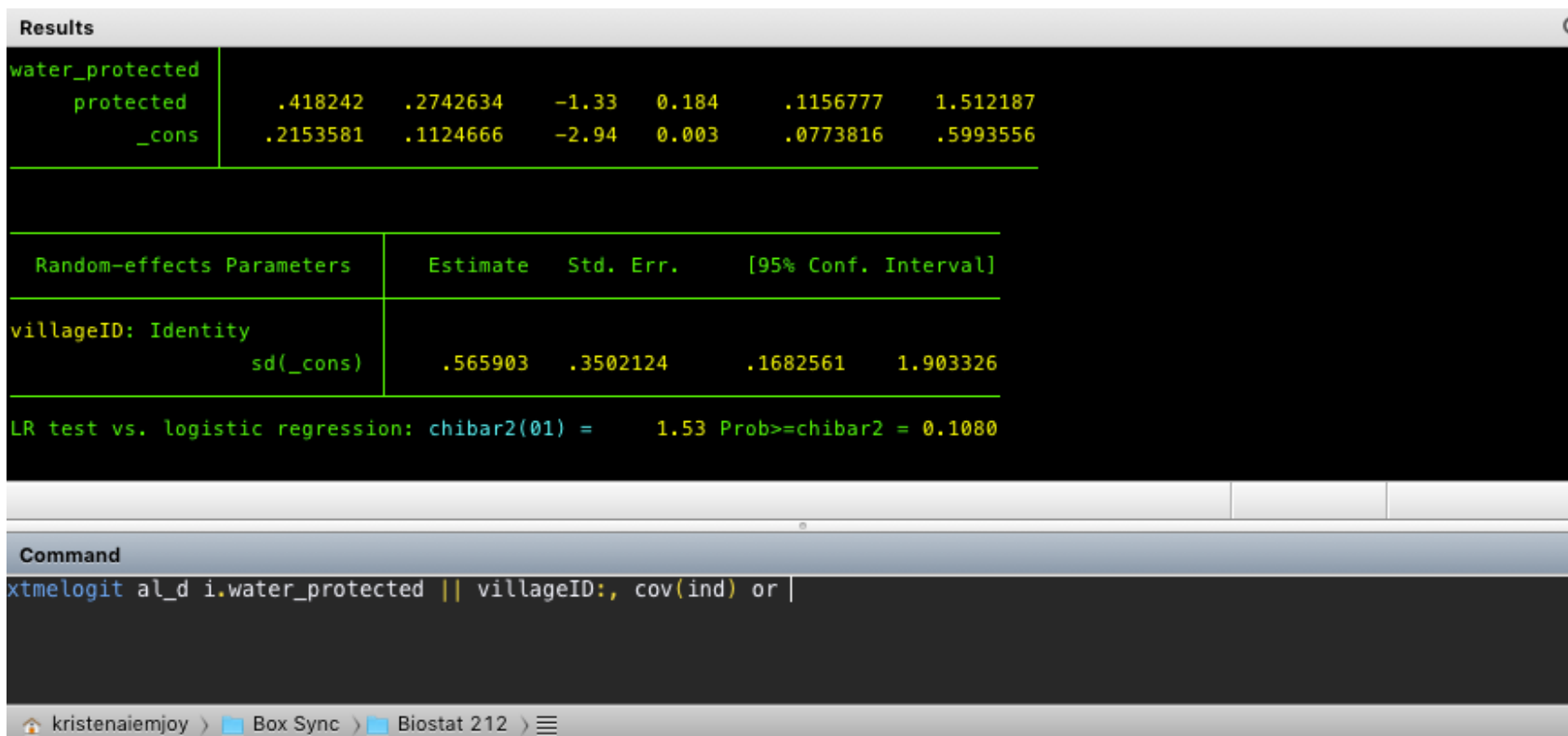
+



Display preferences

◇ Drop down menu

- STATA < General preferences < Syntax Highlighting < Color scheme
- STATA < General preferences < Windows < Results < Color scheme



The screenshot shows the STATA Results window with a dark background and yellow text. The output includes a table of coefficients for the fixed effects, a table for the random effects (villageID), and a likelihood ratio test result. The command window at the bottom shows the command used to run the regression.

```
Results
```

water_protected						
protected	.418242	.2742634	-1.33	0.184	.1156777	1.512187
_cons	.2153581	.1124666	-2.94	0.003	.0773816	.5993556

Random-effects Parameters	Estimate	Std. Err.	[95% Conf. Interval]
villageID: Identity			
sd(_cons)	.565903	.3502124	.1682561 1.903326

LR test vs. logistic regression: chibar2(01) = 1.53 Prob>=chibar2 = 0.1080


```
Command
```

```
xtmelogit al_d i.water_protected || villageID:, cov(ind) or |
```

kristenaiemjoy > Box Sync > Biostat 212 > ≡

Immediate commands

- ◇ An immediate command does not require stored data to operate
 1. Calculator (display)
 - Random number generator
 2. Sample size calculator (power) **[mini-lecture today]**
 3. Statistical/epidemiological immediate commands
 - (end in the letter i)

Immediate commands: Calculator

- ◇ Stata can work as a calculator using the `display` command

```
display 3+5  
8
```

```
display (54.3-21)*83  
2763.9
```

```
display 23*8  
184
```

```
display 17^3  
4913
```

```
display 3+5  
8
```

```
display sqrt(156)  
12.489996
```

Mathematical operators in STATA

English

Stata

Add

+

Subtract

-

Multiply

*

Divide

/

To the power of

^

Natural log of

`ln(expression)`

Base 10 log of

`log10(expression)`

Square Root

`sqrt(#)`

Immediate commands: random number generator

◇ To generate a random number between [a, b]

- `display #a + (#b-#a)*runiform()`

◇ To generate a random integer between [a, b]

- `display #a + int((#b-#a+1)*runiform())`

QUIZ 1

- ◇ Square Root of 457 _____
- ◇ $(10,875 * 456) / (7^2)$ _____
- ◇ Natural log of 342 _____
- ◇ Random Number between 1 and 20 _____
 - `display a + int((b-a+1)*runiform())`

Immediate commands: Statistics

◇ Examples:

- `cii` confidence interval calculator

- `cii #observations #mean #standard deviation`

- `ttesti` T test calculator

- 1 sample:

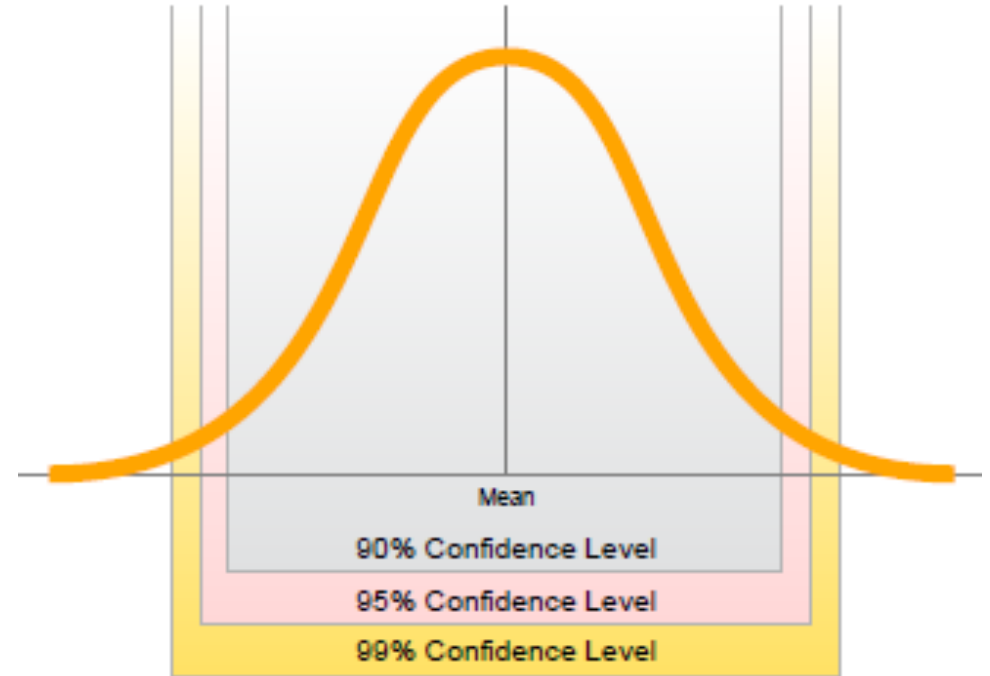
- `ttesti #obs # reported mean #standard deviation #hypothesized mean`

- 2 sample:

- `ttesti #obs1 #mean1 #sd1 #obs2 #mean2 #sd2`

**Preview: confidence intervals

- ◇ Over many repetitions of the study (with different samples) it is expected that 95% of the 95% confidence intervals will include the true (population) value.



QUIZ 2

Abstract

Background: Despite high morbidity and mortality, the laboratory diagnosis of gastrointestinal infections is largely neglected in tropical African settings. This study aims to apply the Luminex multiplex PCR assay for the diagnosis of gastrointestinal pathogens in rural Ghana to evaluate its usefulness as a routine method.

Methods: A case-control study was conducted at the Agogo Presbyterian Hospital in Ghana. Stool samples were collected from children below 6 years of age with (cases) and without (controls) diarrhoea. Samples were screened for 15 different diarrhoeal pathogens by the Luminex xTAG GPP assay and associations between diarrhoea and gastrointestinal infections and fractions attributable to diarrhea (AF) were determined.

Results: The Luminex PCR assay identified organisms in 96.6 % ($n = 428$) of 443 cases and in 92.5 % ($n = 221$) of 239 selected controls. A mean of 2.5 (standard deviation [SD]: ± 1.3) and 2.3 (SD: ± 1.3) organisms per sample were detected in cases and controls respectively. An association with diarrhoea was found for rotavirus (adjusted odds ratio [aOR] = 7.2; 95 % confidence interval [CI]: 2.9–18.1), norovirus (aOR = 2.7; 95 % CI: 1.4–5.3) and *Shigella* spp. (aOR = 1.7; 95 % CI: 1.2–2.4) with respective AFs of 12.5 % (95 % CI: 9.6–15.3), 7.9 % (95 % CI: 3.8–11.7) and 16.9 % (95 % CI: 6.9–25.9).

Conclusion: The high proportion of pathogen-positive stool samples with a high number of co-infections in cases and controls suggests a substantial amount of transient or colonizing microorganisms for which treatment is not necessarily implicated. The use of sequential diagnostic algorithms with pathogen specific or quantitative PCRs might be most appropriate for diagnosing gastrointestinal infections.

Keywords: Ghana, West Africa, Multiplex Polymerase Chain Reaction, Gastrointestinal infections

1. Calculate the 95% confidence interval around the mean number of diarrhea pathogens per stool sample in cases

2. Calculate the 95% confidence interval around the mean number of diarrhea pathogens per stool sample in controls

`cii #observations #mean #standard deviation`

Is the mean number of diarrhea pathogens different between cases and controls?

```
. ttesti 443 2.5 1.3 239 2.3 1.3
```

Two-sample t test with equal variances

	Obs	Mean	Std. Err.	Std. Dev.	[95% Conf. Interval]	
x	443	2.5	.0617649	1.3	2.378611	2.621389
y	239	2.3	.08409	1.3	2.134344	2.465656
combined	682	2.429912	.0498772	1.30255	2.33198	2.527844
diff		.2	.1043361		-.0048597	.4048597

diff = mean(x) - mean(y)

t = 1.9169

Ho: diff = 0

degrees of freedom = 680

Ha: diff < 0

Pr(T < t) = 0.9722

Ha: diff != 0

Pr(|T| > |t|) = 0.0557

Ha: diff > 0

Pr(T > t) = 0.0278

Immediate commands: Epidemiology

◇ Examples:

- `iri` incidence rate calculator
 - `iri #exposed_cases #unexposed_cases #exposed_time #unexposed_time`
- `cci` odds ratio calculator (**case-control** study)
 - `cci #exposed_cases #unexposed_cases #exposed_controls #unexposed_controls`
- `csi` risk/ risk ratio calculator (**cohort** study)
 - `cci #exposed_cases #unexposed_cases #exposed_not_cases #unexposed_not_cases`

**Preview: odds ratio

		Disease	
		Yes	No
Exposure	Yes	a	b
	No	c	d

$$\mathbf{OR} = \frac{\frac{a}{b}}{\frac{c}{d}}$$

For case-control studies

QUIZ 3

		Diarrhea	
		Cases	Controls
Rotavirus	+	64	5
	-	379	234

1. Calculate the odds ratio

```
cci #cases_(+)rotavirus #cases_(-)rotavirus #controls_(+)rotavirus #controls_(-)rotavirus
```

```
. cci 64 379 5 234
```

	Exposed	Unexposed	Total	Proportion Exposed
Cases	64	379	443	0.1445
Controls	5	234	239	0.0209
Total	69	613	682	0.1012
	Point estimate		[95% Conf. Interval]	
Odds ratio	7.902902		3.144012	25.4793 (exact)
Attr. frac. ex.	.8734642		.681935	.9607525 (exact)
Attr. frac. pop	.126189			

```
chi2(1) = 26.06 Pr>chi2 = 0.0000
```

Table 2 Crude odds ratio (OR), age-adjusted OR (aOR) on associations between detected pathogens and diarrhoea, sorted by number of cases

Pathogen ^a	Cases (n = 443)	Controls (n = 239)	Negative PCR-result	Crude OR	aOR
<i>Escherichia coli</i> LT/ST (ETEC)	247 (55.8)	144 (60.3)	291	0.8 (0.6–1.2)	0.9 (0.6–1.2)
<i>Giardia lamblia</i>	228 (51.5)	125 (52.3)	329	0.9 (0.7–1.3)	1.0 (0.7–1.3)
<i>Shigella</i> spp.	185 (41.8)	76 (32.6)	421	1.5 (1.1–2.2)	1.7 (1.2–2.4)
<i>Campylobacter</i> spp.	153 (34.5)	78 (32.6)	451	1.1 (0.8–1.5)	1.1 (0.8–1.6)
Rotavirus	64 (14.5)	5 (2.1)	613	7.9 (3.1–25.5)	7.2 (2.9–18.1)
<i>Escherichia coli</i> O157	59 (13.3)	37 (15.5)	586	0.7 (0.5–1.2)	0.8 (0.5–1.2)
Norovirus	55 (12.4)	11 (4.5)	616	2.9 (1.5–6.3)	2.7 (1.4–5.3)
<i>Salmonella</i> spp.	53 (12.0)	37 (15.5)	592	0.8 (0.5–1.3)	0.8 (0.5–1.3)
<i>Cryptosporidium</i> spp.	33 (7.5)	12 (5.0)	637	1.5 (0.7–3.3)	1.4 (0.7–2.9)
Adenovirus (41/41)	17 (3.8)	14 (5.9)	651	0.6 (0.3–1.4)	0.6 (0.3–1.3)
<i>Escherichia coli</i> STEC	9 (3.7)	12 (2.7)	661	0.7 (0.3–1.9)	0.7 (0.3–1.8)
<i>Clostridium difficile</i> toxin A,B	2 (0.5)	2 (0.8)	678	-	-
<i>Entamoeba histolytica</i>	1 (0.2)	2 (0.8)	679	-	-

^a*Vibrio cholerae* and *Yersinia enterocolitica* were not detected in any sample
OR odds ratio, aOR age adjusted odds ratio